Brown Spot Disease Severity Level Detection using Binary-RGB Image Masking

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Abstract—Agriculture is known as one of the main factor for a growth of a country. Paddy plantation is the most widely planted crop in Malaysia. The rice produced is the main food source to Malaysian's people and source of income to this country as well. However, a disease known as Brown Spot (BS) attacks the paddy plants and threats their quality. This disease caused by bipolar fungus, which represent by the development of an oval, dark brown to purplish-brown spot on leaf. This disease observed as among the hazardous disease that may result in degradation of paddy production. Brown Spot disease could spread through airborne spores from plant to plant on the field. In this research, a system that could help people, especially farmers, to detect the disease at early stage is developed. The real image capture at paddy field is processed in the MATLAB software with image enhancement, background removal as well as binary and RGB image masking process. To determine the Brown Spot area, pixel intensity between the infected and noninfected areas is calculated. The severity level table developed by Horsfall and Heuberger is then used as reference to classify the severity level of Brown Spot disease. A GUI is created to detect the Brown Spot disease automatically. From the study conducted, the accuracy of Brown Spot detection is approximately 89% accurate compared to manual evaluation by plant pathology.

Keywords—Brown spot; image enhancement; binary image; RGB image; masking process

I. INTRODUCTION

Over many years, Malaysia has grown-up its economy through agriculture sector. Agriculture has a significant economic contribution [1], with paddy being the third most widely cultivated crop. Rice produced by paddy, is the main source Malaysian and gain profit to the country [2]. Thus, a lot of efforts have been done to ensure a high quality and quantity production of this crop [3].

Farmers need to take a good care of their paddy growth in order to ensure the production of a good rice quality. However, paddy is very vulnerable as it often threatened by various pest and disease such as leaf folders, stem borer, plant hopper, Brown Spot (BS) disease [4], Bacterial Leaf Blight (BLB) diseases and Leaf Blast (LB) diseases [5].

Brown Spot (BS) disease (see Fig. 1) is caused by the bipolar fungus, making development of an oval, dark brown to purplish-brown spots on leaf. Brown Spot is considered as one of the most severe conditions [6] of paddy plants and can affect paddy leaves by as much as 50 to 90 per cent of the yield

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product. Brown Spot can quickly spread by airborne spores from plant to plant in the field. It occurs when the contaminated seeds are sown at a prevailing low temperature of 18-22°C [7] [8].

Brown Spot disease is a fungal disease that affects the development of rice plants, killing young seeds and reduces their quality. This disease may manifest itself at any stage of crop development [9] but is most severe at full crop maturation. Brown Spot disease is easily identifiable by its characteristic oval to circular form and size of a sesame seed on the surface of the paddy leaf. Typically, the colour is yellow-brown with dark brown patches [10].



Fig. 1. Brown Spot Disease on Paddy Plant.

II. LITERATURE REVIEW

Plant diseases provide a broad scientific field of study in agriculture and emphasize the biological features of diseases. Today, detecting plant diseases is challenging and requires particular care. Fungal infections, bacteria, viruses, and nematodes produce spots on leaves or stems, brown or black lesions, death of lower leaves, yellowing of lower leaves, and black specks [11]. Each disease has its technique of prevention. Standard techniques used include cultural tradition [12], disease resistance cultivars, and chemical usage.

Previously, researcher in [13-14] has developed a system that can detect disease on leaf automatically by using image processing techniques. This, in turn, it allows farmers to recognize the diseases at an early stage and provides valuable knowledge to monitor the crop condition. (IJACSA) International Journal of Advanced Computer Science and Applications, Vol. 12, No. 9, 2021

Image acquisition, image pre-processing, image segmentation, feature extraction, and classification are the image processing stages required for Brown Spot disease diagnosis used in [15-17]. These processes are done on the captured image of infected plants. The diagnosis of plant diseases is usually based on the presence of different colors [18], shapes [19] and abnormalities on plants leaves [20].

The present technique on detecting plant diseases is via professional observation using their bare eyes [21]. To do so, a big team of specialists is required, which is very expensive especially when the number of farms is enormous. At the same time, the farmers do not have enough facilities [22] to do so. In addition, because of this, the expense of consulting experts is often high and time consuming.

In [23], the researcher discusses the detection of two diseases known as Leaf Spot and Leaf Blotch. The procedure used is by divide the process into several stages using image processing techniques. The first step was to convert RGB image to HSI colour space, which retains just the hue component for further processing. The disease part on the leaf is then extracted using K-Means segmentation. After that, the resulting image is analysed for feature extraction using GLCM texture analysis. The output is then used to train and classify a multi-class SVM classifier.

Colour is always the most important element in image processing [24] and serves as an important indication for class identification. Digital image processing produces objective colour measures that are very useful for early lesion detection. The pixel in a colour image is often represented in RGB space, where the colour of each pixel is defined as a triplet (R, G, B), where R, G, and B correspond to Red, Green, and Blue, respectively [25]. The colours in the defect area of the images were analysed to determine the stage of the disease. At this stage, diseases can be categorised into their level according to their condition, from early to the worst stages [2] [26].

Based on these previous researches, it can be seen that the image processing has widely being used in detecting the paddy diseases. Hence, this study is conducted to propose another enhanced technique named Binary-RGB image masking technique to detect Brown Spot disease.

III. METHODOLOGY

The Brown Spot (BS) disease detection start with image acquisition, followed by image enhancement using histogram equalization, background removal, image masking, obtaining the pixel value of segmented RGB image and masked image and determination of Brown Spot Disease based on Disease Severity Scale developed by Horsfall and Heuberger [27] (refer Fig. 2).

The process of the analysis begins with taking pictures of paddy plants that are infected by Brown Spot disease at paddy plot in Malaysian Agricultural Research and Development Institute (MARDI) Seberang Perai using a 24 Megapixel camera with an Optical Image Stabilizer (OIS). The process begins with taking pictures of paddy leaves with a distance of approximately 30 cm.

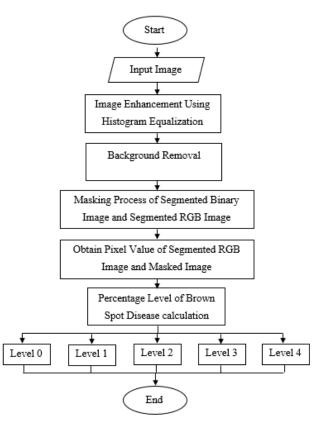


Fig. 2. Flowchart of the Brown Spot Detection Technique.

The quality of the images is then enhanced by using histogram equalization technique. It followed with background removal process to eliminate the similar colour characteristics of the image background with the disease that infected the leaves (see Fig. 3).



Fig. 3. (a) Original Image (b) Enhanced Image by using Histogram Equalization (c) Binary Image for Background Removal (d) RGB Image.

Then, the Brown Spot disease on a leaf is detected by using a masking concept. The procedure starts with masking the segmented RGB image with a segmented binary image. The resulted masked image will show the presence of Brown Spot on the image sample.

As can be seen in Fig. 3(d), the background of an image is successfully removed, leaving the Region of Interest (ROI) with Brown Spot spotted on the leaf. The Brown Spot disease spot are detected after a masking process is applied. To get the pixel of Brown Spot disease, all pixel except black pixel value from a segmented RGB image are then subtracted with the non-black pixel value form by masked image. The resulting value shows the affected area of the Brown Spot disease (see Fig. 4).

This step is continued by obtaining the pixel value for the non-black pixel of the segmented RGB image and masked image. After that, the Brown Spot severity level on the paddy leaf is determined by calculating the lesion and leaf area ratio. The equations used to express this process are as below:

$$S = \frac{Ad}{A1} \tag{1}$$

or S = Pd/P1

Where;

S is severity extent,

Ad is diseases leaf area,

Al is total leaf area,

Pd is total pixel in diseased area,

Pl is total pixel of leaf.



Fig. 4. (a) Original Image (b) Masked Image with Brown Spot Disease Spotted on the Leaves.

 TABLE I.
 DISEASE SEVERITY SCALE DEVELOPED BY HORSFALL AND HEUBERGER [15]

| Level | Severity |
|-------|----------------------------|
| 0 | Apparently infected |
| 1 | 0-25% leaf area infected |
| 2 | 26%-50% leaf area infected |
| 3 | 51%-75% leaf area infected |
| 4 | >75% leaf area infected |

The severity level of the infected area is classified according to the Disease Severity Scale developed by Horsfall and Heuberger as shown in Table I.

IV. ANALYSIS

To get a high quality images, it is essential to pre-process the collected images. In this study, the Histogram Equalization technique is used to improve the image quality. In this technique, there are two parameters used which are 'Radius' and 'Amount' value. The 'Radius' value is used to control the region's edge pixel dimension. The greater the value, the broader is the region around the edge. While a lower value narrows the region around the edge. Besides, the 'Amount' value function is used to increase the contrast of the sharpened pixels. Therefore, with a larger value of 'Amount', the brightness of the image will increase.

As shown in Fig. 5, there are some changes happen in the histograms of the Red, Green, and Blue channels between the original and enhanced images. The x-axis indicates the tonal scales, while the y-axis indicates the number of pixels in an image. If the pixel's value on the X-axis is close to 0, it indicates that the pixel has a darker black colour. While if its value near to 255, that certain pixel is merely get a lighter colour or white.

By using Histogram Equalization technique the most common intensity pixel values are efficiently spreads out the while stretching the image's intensity range. This technique, in particular, is resulting in an improved image intensity as well as image quality of the Region of Interest (ROI), which in this instance is the paddy leaf.

As for example, the peak value of blue channel is at 50, while the green channel is on range of 0-50, and red channel is on the range of 0 to 250. After the Histogram Equalization process, the pixel intensity value is being stretched out to it closes contrast value. As for the pixel value of blue channel, the colour pixel of 50 changes from 8000 to 7000, where the colour pixel of this channel is stretched out to its near intensity range. For green channel, the colour pixel is spreading towards 0 which indicate the image at the certain pixel will become darker. At the red channel, the colour pixel is spreading more towards the 250 where it's resulting to a brighter pixel. This will increases the image contrast when its data is represented by close contrast values.

(2)

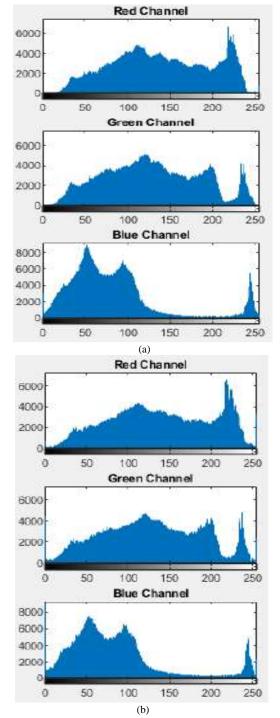


Fig. 5. (a) Histogram for Original Image (b) Histogram for Enhanced Image.

The image background on enhanced image is then removed and keep only the object of interest, which is the paddy leaves. Then, the affected area on the paddy leaf is detected by subtracting the segmented RGB image with the masked image. The output of two different sample can be seen in Fig. 6 and Fig. 7.

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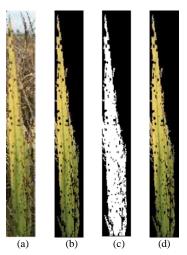


Fig. 6. (a) Original Image (b) Segmented RGB Image (c) Segmented Binary Image (d) Masked Image.



Fig. 7. (a) Original Image (b) Segmented RGB Image (c) Segmented Binary Image (d) Masked Image.

In this technique, it is compulsory to get the total pixels of leaf and the total pixel of Brown Spot affected area first. To get the pixel value of the Brown Spot affected area, the non-black pixel is obtained from masked image while for total pixel of leaf is obtained from subtracting the non-black pixel of masked image and the not black pixel of binary image.

In order to obtain the Region of Interest (ROI), the value of the non-black pixel for the segmented RGB image and segmented binary image is calculated. Here, the Horsfall and Heuberger method [15], is applied to calculate the severity level of the Brown Spot disease on the paddy leaf. The equation for Horsfall and Heuberger method is as follows:

$$S = 100 x \left[\frac{B-K}{B}\right]$$

Where;

B is the non-black pixel of the segmented RGB image,

K is the non-black pixel of the masked image,

For example, for sample image in Fig. 6, the non-black pixel value of segmented RGB image is 64966, while non-black pixel value of masked image is 16728. So, the calculated affected area is;

Affected area

=100×((B-K)/B)

=100*((64966-43922)/64966)

=32.39%

Based on the Disease Severity Scale, if the percentage obtained is in the range of 26%-50%, the sample image is classified as affected with Brown Spot disease at Level 2. Since the sample image as in Figure 6 is calculated to have an affected percentage of 32.39% so, it fall under Brown Spot severity Level 2.

As for image sample in Fig. 7, the non-black pixel value of segmented RGB image is 47501, while non-black pixel value of masked image is 36981. So, the calculated affected area is;

Affected area

=100×((B-K)/B)

=100*((47501-36981)/47501)

=22.15%

So, based on Disease Severity Scale, this sample fall under Brown Spot severity Level 1.

These image samples are also being forwarded to Plant Pathology for a manual evaluation. The manual observation shows that sample image in Fig. 6 was 30% effected by Brown Spot. While for sample image in Fig. 7, it was manually evaluated as 20% effected with Brown Spot.

Table II shows the result of Brown Spot area percentage by using system calculation and the manual evaluation by plant patology on five collected samples. While Table III shows the Brown Spot severity level by using system calculation and the manual evaluation.

As can be seen in Table II, the result of Brown Spot area percentage by using system calculation and manual evalution are sightly different but still tolarable. The precision in detecting the Brown Spot disease by using proposed system is about 89%. Although the percentage area of Brown Spot detected on image sample are different, but the severity level by using system calculation and manual evaluation are the same.

To ease the process so that the layman can use the proposed system, a GUI as shown in Fig. 8 is developed. Here, the user need to load the original image to the system. Then, the system will automatically shows the enhanced image, segmented RGB image, segmented binary image and masked image. At the same time, the non-black pixel value of the RGB and masked image, the percentage of disease severity and its severity level is calculated and the result will be appear in the table of severity level box. The reset button is also available to clear all the input data as preparation to receive a new image sample.

 TABLE II.
 BROWN SPOT AREA PERCENTAGE BY USING SYSTEM

 CALCULATION AND THE MANUAL EVALUATION

| Sample | Calculation on System | Manual Evaluation |
|--------|-----------------------|-------------------|
| 1 | 39.48% | 35% |
| 2 | 32.65% | 30% |
| 3 | 57.58% | 55% |
| 4 | 21.30% | 20% |
| 5 | 70.96% | 60% |

 TABLE III.
 BROWN SPOT SEVERITY LEVEL BY USING SYSTEM

 CALCULATION AND THE MANUAL EVALUATION

| Sample | Calculation on System | Manual Evaluation |
|--------|-----------------------|-------------------|
| 1 | Level 2 | Level 2 |
| 2 | Level 2 | Level 2 |
| 3 | Level 3 | Level 3 |
| 4 | Level 1 | Level 1 |
| 5 | Level 3 | Level 3 |

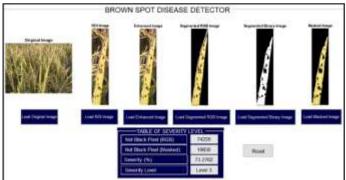


Fig. 8. GUI of the Proposed System.

V. CONCLUSION

This project is intended to help people, especially farmers who work in the agriculture sector. Instead of using too many pesticides that could affect people's health, early detection of the disease should help get rid of the disease before it gets worse and endanger the paddy's quality. Therefore, by using the proposed technique, it will help the farmer to classify the severity level of Brown Spot disease and it may assist the farmers in the early detection of the Brown Spot disease before it spread widely in the paddy field.

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REFERENCES

- [1] S. C. Omar, A. Shaharudin, and S. A. Tumin, The Status of the Paddy and Rice Industry in Malaysia. Khazanah Research Institute, 2019.
- [2] S. Mutalib, M. H. Abdullah, S. Abdul-Rahman, and Z. A.Aziz, "A Brief Study on Paddy Applications with Image Processing and Proposed

Architecture," in 2016 IEEE Conference on Systems, Process and Control (ICSPC 2016), 2016, no. December, pp. 124–129, doi: 10.1109/SPC.2016.7920716.

- [3] M. M. M. Najim, T. S. Lee, M. A. Haque, and M. Esham, "Sustainability of rice production: a Malaysian perspective," J. Agric. Sci., vol. 3, pp. 1– 12, 2007, doi: 10.4038/jas.v3i1.8138.
- [4] B. S. Anami, N. N. Malvade, and S. Palaiah, "Classification of yield affecting biotic and abiotic paddy crop stresses using field images," Inf. Process. Agric., vol. 7, no. 2, pp. 272–285, 2020.
- [5] P. A. C. Ooi, "Common insect pests of rice and their natural biological control," Agric. Sci. J., vol. 1, no. 1, pp. 49–59, 2015.
- [6] R. P. Narmadha, "Detection and Measurement of Paddy Leaf Disease Symptoms using Image Processing," pp. 26–29, 2017.
- [7] L. Aryal, G. Bhattarai, A. Subedi, M. Subedi, B. Subedi, and G. K. Shah, "Response of Rice Varieties to Brown Spot Disease of Rice at Paklihawa , Rupandehi," Glob. Institute Res. Educ., vol. 5, no. 2, pp. 50–54, 2016.
- [8] Z. Mohamed, R. Terano, M. N. Shamsudin, and I. A. Latif, "Paddy Farmers' Sustainability Practices in Granary Areas in Malaysia," Resources, vol. 5, pp. 1–11, 2016, doi: 10.3390/resources5020017.
- [9] A. Sharma, D. Satish, and S. Sharma, "Indian major basmati paddy seed varieties images dataset," Data Br., vol. 33, p. 106460, 2020.
- [10] I. Ihsan, E. W. Hidayat, and A. Rahmatulloh, "Identification of Bacterial Leaf Blight and Brown Spot Disease In Rice Plants With Image Processing Approach," J. Ilm. Tek. Elektro Komput. dan Inform., vol. 5, no. 2, pp. 59–67, 2019, doi: 10.26555/jiteki.v5i2.14136.
- [11] N. S. A. M. Taujuddin et al., "Detection of Plant Disease on Leaves using Blobs Detection and Statistical Analysis," Int. J. Adv. Comput. Sci. Appl., vol. 11, no. 8, pp. 407–411, 2020.
- [12] R. G. De Luna, E. P. Dadios, and A. A. Bandala, "Automated Image Capturing System for Deep Learning-based Tomato Plant Leaf Disease Detection and Recognition," in IEEE Region 10 Conference, Proceedings of TENCON, 2018, vol. 2018-Octob, no. October, pp. 1414–1419, doi: 10.1109/TENCON.2018.8650088.
- [13] R. S. Indumathi, N.Saagari, V.Thejuswini, "Leaf Disease Detection And Fertilizer Suggestion," in Proceeding of International Conference on Systems Computation Automation and Networking, 2019, pp. 1–7.
- [14] S. Kn, M. Suresha, and H. N. T, "A Novel Segmentation and Identification of Diseases in Paddy Leaves Using Color Image Fusion Technique," pp. 17–22, 2021.
- [15] S. Ramesh and D. Vydeki, "Recognition and classification of paddy leaf diseases using Optimized Deep Neural network with Jaya algorithm," Inf. Process. Agric., vol. 7, no. 2, pp. 249–260, 2020, doi: 10.1016/j.inpa.2019.09.002.

- [16] M. V. Overbeek, "Identification of Maize Leaf Diseases Cause by Fungus with Digital Image Processing (Case Study: Bismarak Village Kupang District – East Nusa Tenggara)," in 5th International Conference on New Media Studies, 2019, pp. 125–128.
- [17] S. Ramesh and D. Vydeki, "Application of machine learning in detection of blast disease in South Indian rice crops," J. Phytol., vol. 11, pp. 31–37, 2019.
- [18] G. Dhingra, V. Kumar, and H. D. Joshi, "Study of digital image processing techniques for leaf disease detection and classification," Multimed. Tools Appl., pp. 19951–20000, 2018.
- [19] M. M. Tin, M. M. Khin, S. S. Hlaing, P. P. Wai, and K. L. Mon, "Leaves Disease and Damage Rate Classification based on Features," pp. 419– 420, 2021.
- [20] T. Islam, M. Sah, S. Baral, and R. Roychoudhury, "A Faster Technique on Rice Disease Detectionusing Image Processing of Affected Area in Agro-Field," Proc. Int. Conf. Inven. Commun. Comput. Technol. ICICCT 2018, no. Icicct, pp. 62–66, 2018, doi: 10.1109/ICICCT.2018.8473322.
- [21] N. N. Kurniawati, S. N. H. S. Abdullah, S. Abdullah, and S. Abdullah, "Texture analysis for diagnosing paddy disease," Proc. 2009 Int. Conf. Electr. Eng. Informatics, ICEEI 2009, vol. 1, no. August, pp. 23–27, 2009, doi: 10.1109/ICEEI.2009.5254824.
- [22] V. Singh, Varsha, and A. K. Misra, "Detection of unhealthy region of plant leaves using image processing and genetic algorithm," in 2015 International Conference on Advances in Computer Engineering and Applications(ICACEA 2015), 2015, pp. 1028–1032, doi: 10.1109/ICACEA.2015.7164858.
- [23] G. Kuricheti and P. Supriya, "Computer Vision Based Turmeric Leaf Disease Detection and Classification: A Step to Smart Agriculture," in Proceedings of the Third International Conference on Trends in Electronics and Informatics (ICOEI 2019), 2019, no. Icoei, pp. 545–549, doi: 10.1109/ICOEI.2019.8862706.
- [24] N. S. A. M. Taujuddin, R. Ibrahim, and S. Sari, "Progressive Pixel-to-Pixel Evaluation to Obtain the Hard and Smooth Region for Image Compression," in Proceedings - International Conference on Intelligent Systems, Modelling and Simulation, ISMS, 2015, Vol. 2015.
- [25] M. V Latte and S. Shidnal, "Multiple Nutrient Deficiency Detection in Paddy Leaf Images using Color and Pattern Analysis," 2016 Int. Conf. Commun. Signal Process, pp. 1247–1250, 2016.
- [26] N. N. Kurniawati, S. N. H. Sheikh Abdullah, S. Abdullah, and S. Abdullah, "Investigation on Image Processing Techniques for Diagnosing Paddy Diseases," in 2009 International Conference of Soft Computing and Pattern Recognition, 2009, pp. 272–277, doi: 10.1109/SoCPaR.2009.62.
- [27] S. B. Patil and S. K. Bodhe, "Leaf Disease Severity Measurement using Image Processing," Int. J. Eng. Technol., vol. 3, no. 5, pp. 297–301, 2011.